Evaluation of Stationary Phases for the Analysis of Volatiles by US EPA Methods 8260 & 524.2.

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Abstract

There are many columns that are currently used for the analysis of volatile environmental pollutants by GC/MS. Most of the stationary phases that are offered have adequate selectivity, bleed and inertness for the most common methods. Problems occur when starting temperatures & oven ramp rates are increased. Laboratories that increase runtimes without carefully checking the mass-spectra of each coeluting analyte may compromise data integrity.

This paper will compare four common stationary phases to a new volatile-mass / spec phase for separation of isomeric pairs using high oven starting temperatures and a rapid final oven ramp rate. Other common analytes that share ions will also be presented.

Introduction

Chromatographic and purge & trap cycle times have been the limiting factor for volatile analysis for many years. With the advent of interfacing two purge & trap concentrators onto one GC/MS, analysts are now interested in columns with sub-tenminute runtimes.

The Rtx-VMS chromatogram shown below spectrally resolves all of the target compounds in under 10 minutes. Faster runtimes are possible by substituting chlorobenzene-D5 with an alternate internal standard. This solution works for the VMS phase but will not work for other phases since the resolution of isomers is compromised. This paper will compare other phases run under these conditions for separation of isomeric pairs.

20m, 0.18 mm ID, 1.00µm Rtx*-VMS (cat.# 49914) Compounds in at 10ppb in 5mL of RO water unless otherwise noted, ketones in at 2.5X Concentrator: Tekmar LSC-3100 Purge and Trap

Trap: Vocarb 3000 (type K)

Purge: 11 min. @ 40mL/min. @ ambient temperature

Dry purge: 1 min. @ 40mL/min.

Desorb preheat:245°C

50. toluene-d8(SMC)

Desorb: 250°C for 2 min., Flow 40mL/min.

Bake: 260°C for 8 min.

Interface: transfer line 0.53mm ID Silcosteel* tubing 1:40 split at injection port. 1mm ID sleeve.

Oven temp.: 50°C (hold 4 min.) to 100°C @ 18°C/min. (hold 0 min.)

to 230°C @ 40°C/min. (hold 3 min.)

Carrier gas: helium @ ~1.0mL/min. constant flow

Adjust dichlorodifluoromethane to a retention time of 1.03 min. @

50«M,29[Degreeered Trademark]»C. Detector: HP 5973 MSD Scan range: 35-300amu

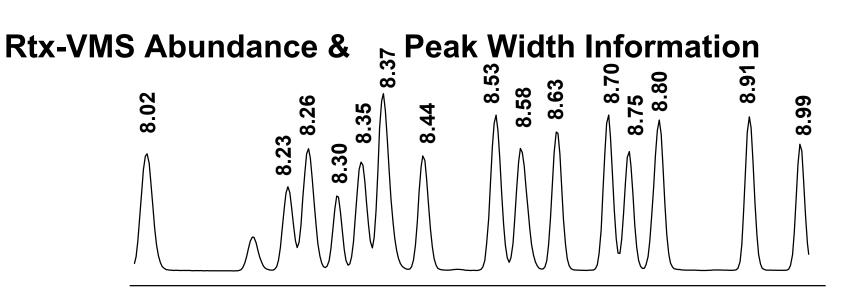
76. *n*-propylbenzene 77. 1,1,2,2-tetrachloroethane 78. 2-chlorotoluene 1. dichlorodifluoromethane 51. toluene 26. ethyl acetate 52. pyridine (250ppb) 2. chloromethane 27. carbon tetrachloride 3. vinyl chloride 28. methyl acrylate 53. tetrachloroethene 29. propargyl alcohol (500ppb) 30. dibromofluoromethane (SMC) 79. 1,3,5-trimethylbenzene 4. bromomethane 54. 4-methyl-2-pentanone 80. 1,2,3-trichloropropane 81. 4-chlorotoluene 5. chloroethane 55. *trans*-1,3-dichloropropene 56. 1,1,2-trichloroethane 6. trichlorofluoromethane 31. tetrahydrofuran 7. ethanol (2500ppb) 32. 1,1,1-trichloroethane 57. ethyl methacrylate 82. tert-butylbenzene 83. pentachloroethane 84. 1,2,4-trimethylbenzene 8. 1,1-dichloroethene 33. 2-butanone 58. dibromochloromethane 9. carbon disulfide (40ppb) 34. 1,1-dichloropropene 59. 1,3-dichloropropane 10. allyl chloride 35. benzene 60. 1,2-dibromoethane 85. sec-butylbenzene 11. methylene chloride 36. pentafluorobenzene (IS) 86. *p*-isopropyltoluene 87. 1,3-dichlorobenzene 61. *n*-butyl acetate 37. *tert*-amyl-methyl ether 38. 1,2-dichloroethane 12. acetone 62. 2-hexanone 63. 2-picoline (250ppb) 64. chlorobenzene-D5 (IS) 13. trans-1,2-dichloroethene 88. 1,4-dichlorobenzene-d4 (IS) 14. methyl tert-butyl ether 39. isobutyl alcohol (500ppb) 89. 1,4-dichlorobenzene 15. *tert*-butyl alcohol (100ppb) 40. isopropyl acetate 65. chlorobenzene 90. n-butylbenzene 16. diisopropyl ether 17. 1,1-dichloroethane 18. acrylonitrile 66. ethylbenzene 67. 1,1,1,2-tetrachloroethane 91. 1,2-dichlorobenzene 41. trichloroethene 92. 1,2-dibromo-3-chloropropane 93. nitrobenzene (250ppb) 42. 1,4-difluorobenzene (SMC) 43. dibromomethane 68. m-xylene 19. vinyl acetate 20. allyl alcohol (250ppb) 44. 1,2-dichloropropane 45. bromodichloromethane 94. hexachlorobutadiene 95. 1,2,4-trichlorobenzene 69. *p*-xylene 70. *o*-xylene 21. ethyl-tert-buyl ether 46. methyl methacrylate 71. stryrene 96. naphthalene 22. *cis*-1,2-dichloroethene 47. n-propyl acetate 72. bromoform 97. 1,2,3-trichlorobenzene 23. 2,2-dichloropropane 48. 2-chloroethanol (2500ppb) 73. isopropylbenzene 49. *cis*-1,3-dichloropropene 74. 4-bromo-1-fluorobenzene (SMC) 24. bromochloromethane

75. bromobenzene

25. chloroform

Rtx-VMS Abundance & Peak Width Information

The EZ-GC® computer program allows up to 100 compounds to be modeled on a single stationary phase under varying GC conditions & column dimensions with an excellent level of both precision and accuracy. Retention time, abundance, and peak width were loaded into EZ-GC®. The abundance and peak width values were determined from the Rtx-VMS chromatogram which was run using an HP5973 GC/MS system (see GC_EV00428 for specific conditions).



			MS	
Rtx-VMS			Abundance	
Compounds Examined	RT (min.)	Peak Wi	(x 1000)	
isopropylbenzene	8.02	0.045	440	
bromobenzene	8.23	0.058	340	
n-propylbenzene	8.26	0.055	460	
1,1,2,2-tetrachloroethane	8.30	0.05	290	
2-chlorotoluene	8.35	0.04	400	
1,3,5-trimethylbenzene	8.37	0.045	440	
1,2,3-trichloropropane	8.37	0.06	180	
4-chlorotoluene	8.44	0.04	400	
tert-butylbenzene	8.53	0.05	550	
1,2,4-trimethylbenzene	8.58	0.055	440	
sec-butylbenzene	8.63	0.045	525	
p-isopropyltoluene	8.70	0.055	545	
1,3-dichlorobenzene	8.75	0.04	460	
1,4-dichlorobenzene	8.80	0.04	475	
n-butylbenzene	8.91	0.055	501	
1,2-dichlorobenzene	8.99	0.04	475	

Peak Widths & Abundance Determined from Purge and Trap Analysis.

Rtx-VMS Model versus Actual Analysis

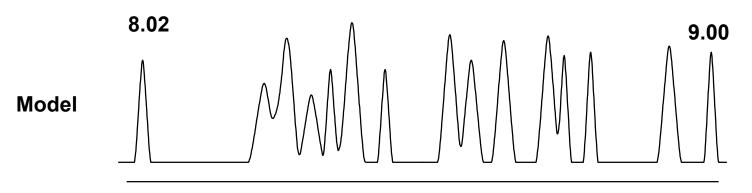
Using retention time indices specific to each of the stationary phases in combination with the abundance values & peak widths values from the VMS phase; model chromatograms were produced. Since column efficiency may change with each stationary phase – only differences in retention time will be examined. The abundance and peak width information was loaded into the program to produce sample chromatograms as a visual aid only.

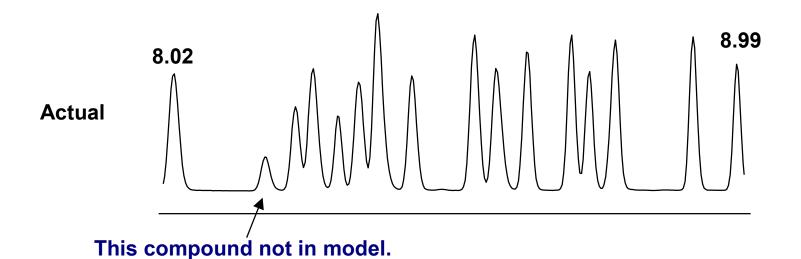
The first step in the column comparison was to determine the effectiveness of EZ-GC® by comparing our known VMS chromatogram with a model. Retention times were collected for these compounds under fast and slow conditions and were entered into EZ-GC®. The application chromatogram was modeled using this fast/slow data. The diagram below shows the difference between the actual chromatogram and the model to be 0.01 minutes.

Rtx-VMS Model versus Actual Analysis

Rtx-VMS 20 m x 0.180 mm x 1.0 μm

50°C (4) @ 18°C/min to 100°C @ 40°C/min to 230°C (1) Flow: ~ 1.0 ml/min

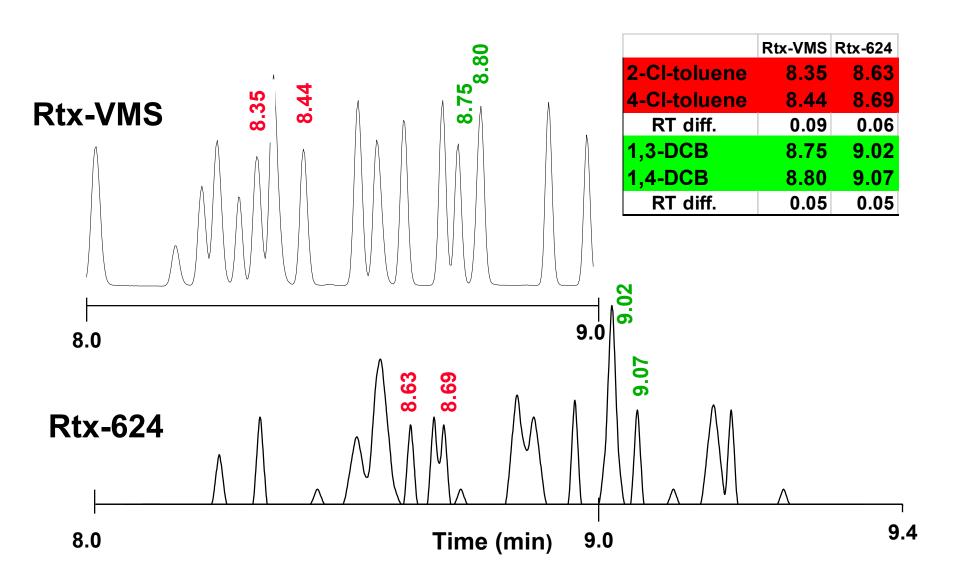




Rtx-VMS Compared to the Rtx-624

The change in retention between the chlorotoluenes modeled on the VMS & 624 phases is 0.03 minute, which in reality, constitutes a significant difference since this data is produced with a 20m x 0.18mm ID column with a 40°C/minute oven ramp rate. A faster final oven ramp rate is possible using the VMS phase since these compounds are well separated. Using a 60m x 0.53mm ID x 3.0df column for a direct comparison the difference between these phases is 0.9 minute. The 624 phase is suitable for fast dual-purge & trap applications. Analytes that share ions and coelute on the 624 phase and are resolved by the VMS include: ether/ethanol, vinyl acetate/ethyl-tert-butyl-ether & t-butyl alcohol/methyl-tert-butyl-ether. Several of these compounds require a lower starting oven temperature (35°C) using the VMS phase which is not shown in this application.

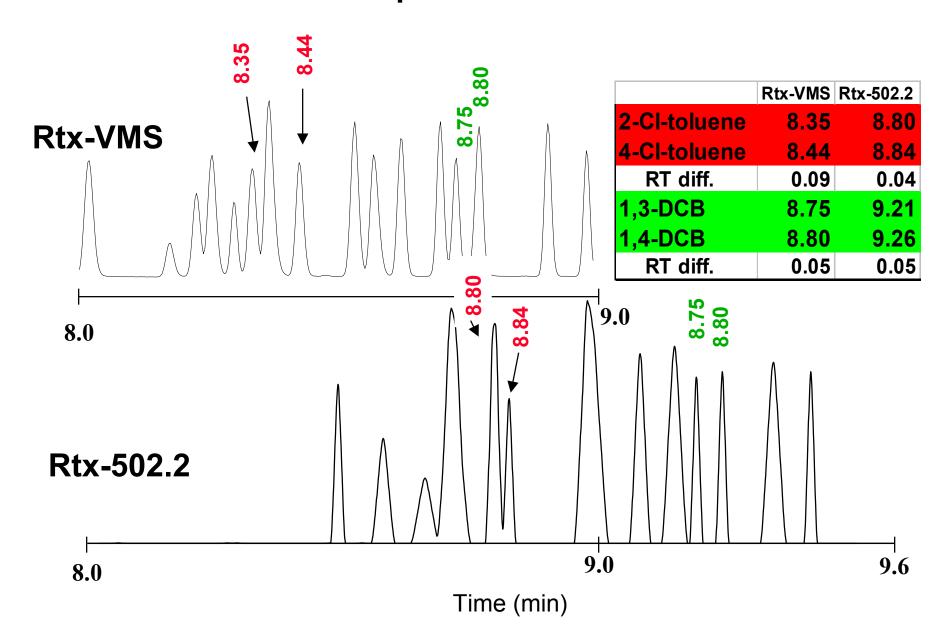
Rtx-VMS Compared to the Rtx-624



Rtx-VMS Compared to the Rtx-502.2

The Rtx-502.2 is a cross-bonded diphenyl-methyl containing polysiloxane, which is particularly resistive to oxidative breakdown and exhibits lower bleed compared to the cyanopropyl-phenyl polysiloxanes (624phase). The disadvantage is the difficult resolution between bromomethane and chloroethane. Although our model indicates partial resolution between the chlorotoluenes under these runconditions, linearity is effected over the concentration range. Slower final oven ramp rates will resolve these isomers, but it will sacrifice runtime.

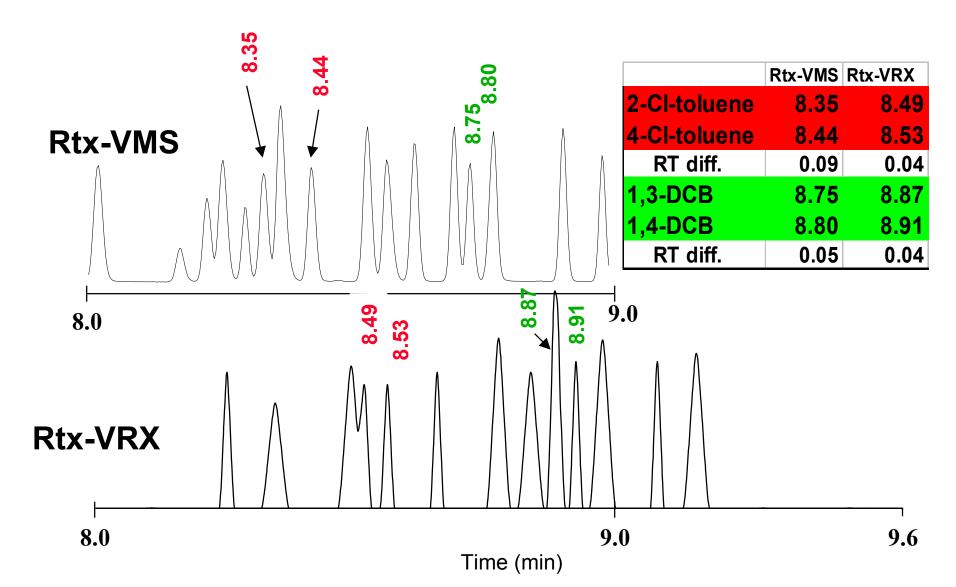
Rtx-VMS Compared to the Rtx-502.2



Rtx-VMS Compared to the Rtx-VRX

The Rtx-VRX incorporates a low percentage of pentafluorobenzylpropyl- polysiloxane, which gives this column similar characteristics to an Rtx-1, only it resolves close eluting GC pairs such as 2,2-dichloropropane & chloroform. It is possible to do fast dual purge and trap applications using this phase as long as the recommended EPA internal standard, chlorobenzene-d5 is substituted with another compound. As shown in the direct comparison the Rtx-VMS does out-perform the VRX stationary phase for this application.

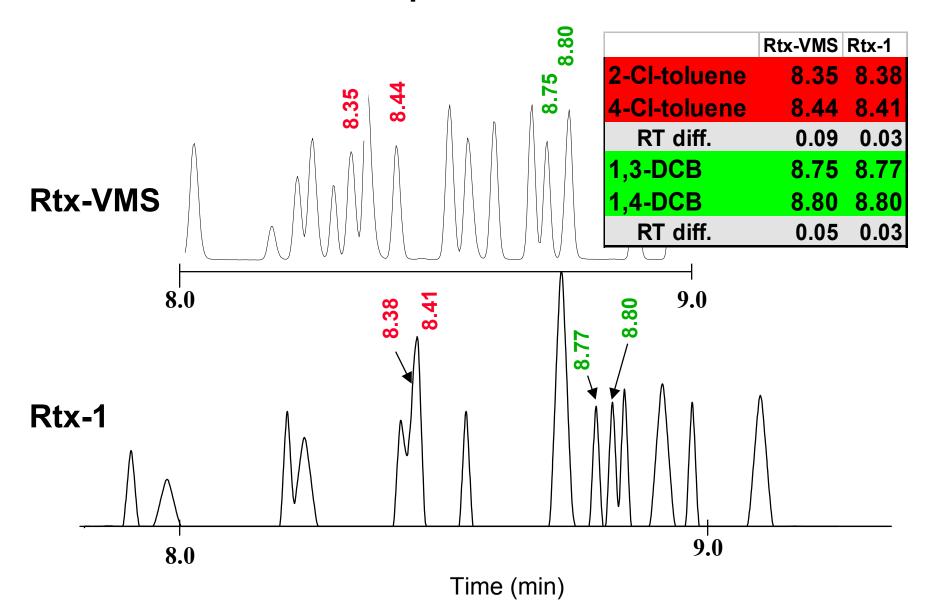
Rtx-VMS Compared to the Rtx-VRX



Rtx-VMS Compared to the Rtx-1

Although the 100% dimethyl-polysiloxane (Rtx-1) polymer is unchallenged for air analysis, this phase does not perform well for fast purge and trap analysis. Like the VRX phase chlorobenzene-d5 & 1,1,1,2-tetrachloroethane are poorly resolved as are the isomers shown.

Rtx-VMS Compared to the Rtx-1



The VMS phase has the best resolution of isomeric pairs and other target compounds for US EPA Method 8260B.

Rtx-VMS – Best Resolution of Isomeric Pairs.

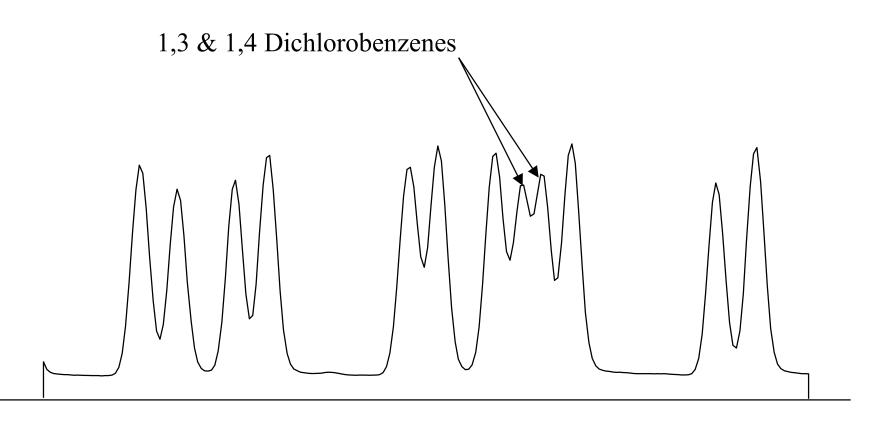
	Shared					
<u>COMPOUND</u>	lons	Rtx-VMS	Rtx-624	Rtx-502.2	Rtx-VRX	Rtx-1
Chlorobenzene-d5	117	7.34	7.62	7.90	7.34	7.25
1,1,1,2-tetrachloroethane	117	7.42	7.70	7.93	7.39	7.24
RT diff.		0.08	0.08	0.07	0.04	0.01
2-chlorotoluene	126,91	8.35	8.63	8.80	8.49	8.38
4-chlorotoluene	126,91	8.44	8.69	8.84	8.53	8.41
RT diff.		0.09	0.06	0.04	0.04	0.03
1,3-dichlorobenzene	146	8.75	9.02	9.21	8.87	8.77
1,4-dichlrorobenzene	146	8.80	9.07	9.26	8.91	8.80
RT diff.		0.05	0.05	0.05	0.04	0.03

A Candidate for Volatile Applications Fails for Resolution of the DCBs.

One of our attempts to build a column suitable for fast volatile analysis met all of the design criteria except for the resolution between the dichlorobenzenes.

Experimental Fluorinated Phase

A Candidate for Volatile Applications Fails for Resolution of the DCBs.

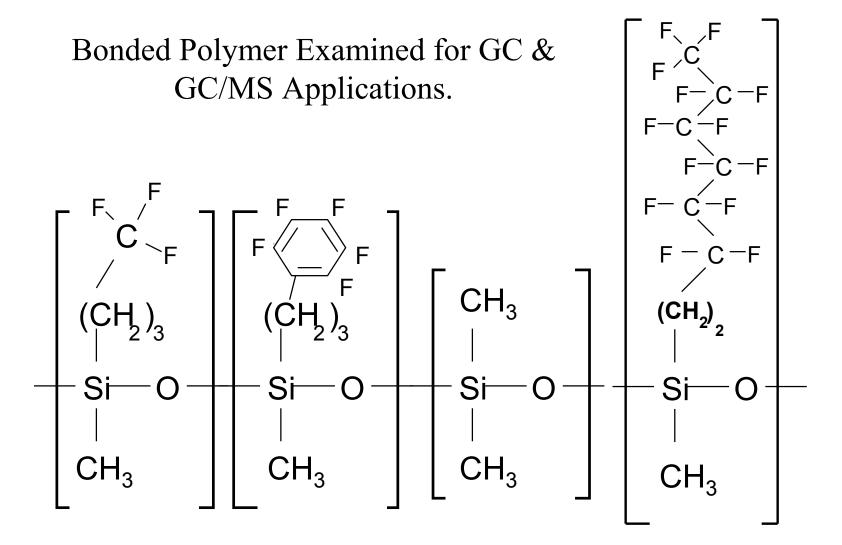


Experimental Fluorinated Phase

Bonded Polymer Examined for Volatile Applications.

This phase was built based on predictions of separation. The separation criteria was met on all accounts except for the dichlorobenzenes. The first step in designing a new column is to start with functionalities that the target analytes are soluble in. The better the solubility between the analyte and the phase the better the chances of resolving these compounds.

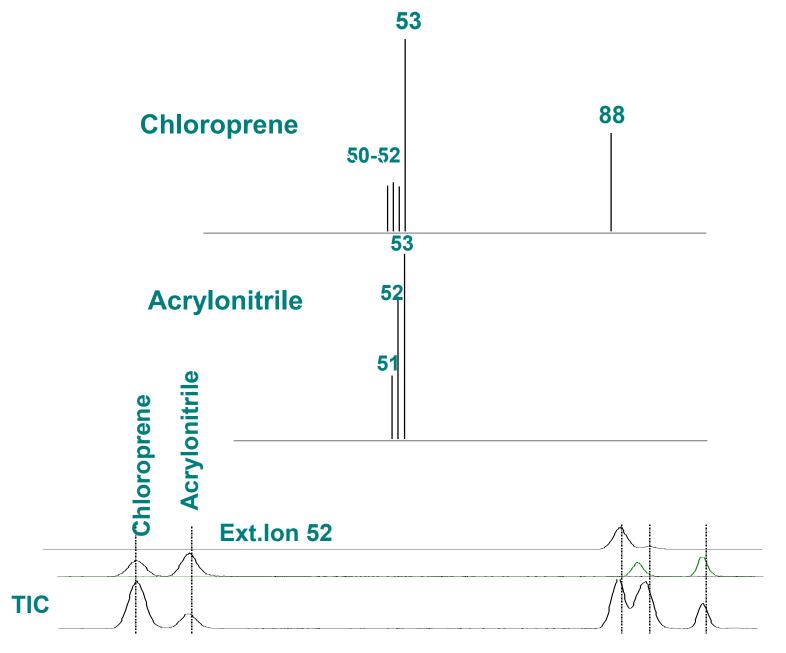
Experimental Fluorinated Phase



Rtx-VMS Column Meets Design Criteria

The successful polymer known as the VMS phase not only met the resolution criteria for the isomeric pairs but also exceeded separations of critical pairs. The percentage of functionality was adjusted to resolve chloroprene and acrylonitrile.

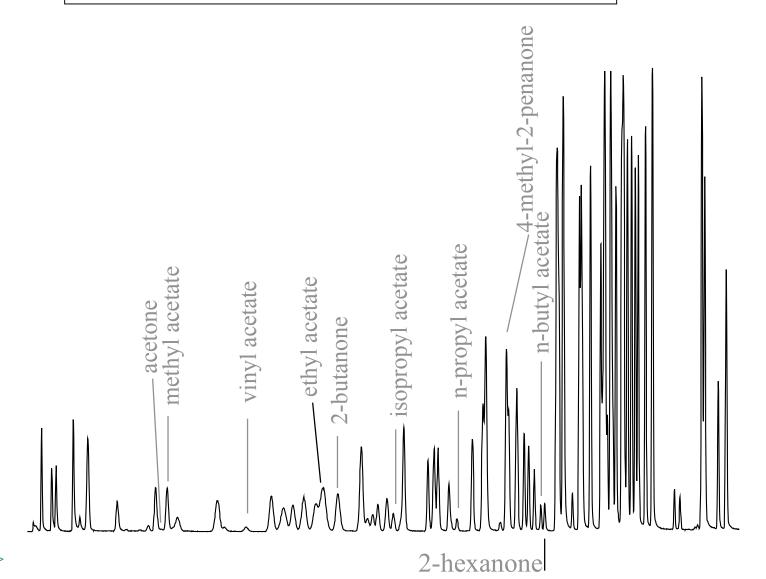
Rtx®-VMS Critical Pairs with Common Ions



Rtx-VMS Resolution of Acetates & Ketones.

Since the acetates and ketones share the spectral ion 43 its important that these analytes are resolve. The diagram shows the elution of the most commonly analyzed acetates and ketones.

Acetates & Ketones Share ion 43



Rtx-VMS Designed for EPA Method 8260B

The Rtx-VMS column was designed to address the increasing number of analytes listed in EPA Method 8260B, and also is a good choice for separating compounds listed in EPA Method 524.2, revision IV. The major difference between the Rtx-VMS phase and others such as the "624," "502.2," and "VRX" is its overall selectivity and the distance between isomeric pairs like 2/4-chlorotoluene. A faster final oven ramp rate is possible because these compounds elute farther apart on the Rtx-VMS phase, preventing a partial coelutions that would interfere with quantitation. Using the EPA-suggested surrogates the analysis time is under 10 minutes making this the clear choice for fast GC applications.

For More Information...

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